

## E. Description of Specific Growth Rate – Unstructured Models

### 1. Limiting Substrate Concentration

That is,  $\mu = f(S)$

**Empirical** Models are Used:

a) Monod  $\mu = \frac{\mu_{MAX} S}{K_S + S}$   $K_S = \text{“Monod Constant”}$   
(typically very small)

So,  $\mu \approx \mu_{MAX}$

For multiple substrates...  $\mu = \frac{\mu_{MAX} S}{K_S + S} \frac{O}{K_O + O}$

b) Modified Monod

$$\mu = \frac{\mu_{MAX} S}{K_S + K_{S_0} S_0 + S}$$

$S_0$  = Initial Substrate  
Concentration

c) Blackman

$$\mu = \mu_{MAX}$$

If  $S \geq 2K_S$

$$\mu = \frac{\mu_{MAX} S}{2K_S}$$

If  $S < 2K_S$

No transition between 0<sup>th</sup> and 1<sup>st</sup> order

d) Tessier  $\mu = \mu_{MAX} (1 - e^{-KS})$

e) Moser  $\mu = \frac{\mu_{MAX} S^N}{K_S + S^N}$  Same as Monod when N=1

f) Contois  $\mu = \frac{\mu_{MAX} S}{K_{SX}X + S}$  Predicts  $\mu \propto \frac{1}{X}$  at low S

Predicts  $\mu \rightarrow 0$  at high X

## Comments

Monod most commonly used model

Blackman is discontinuous

Contois often preferred for wastewater treatment.  
Also, unlike other models in a continuous process it allows for effluent concentration ( $S_{OUT}$ ) to be dependent of influent concentration ( $S_{IN}$ )

Contois and Tessier show most gradual decrease in specific growth rate at low substrate concentrations

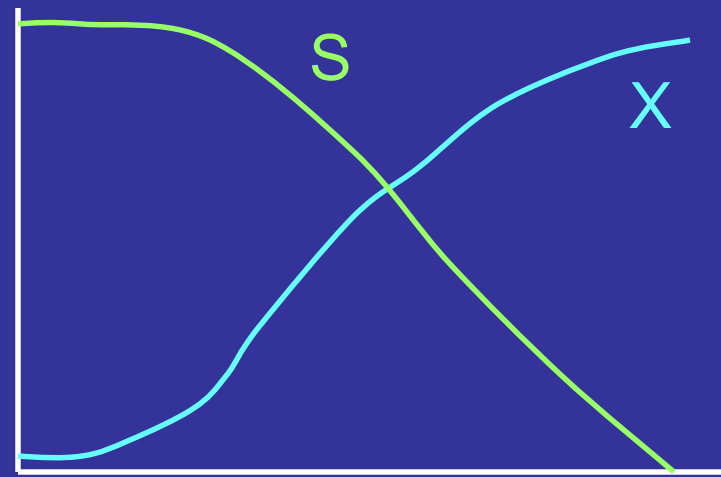
Moser most general

All models are very similar

## How are parameters determined?

1) Measure S and X as a function of time

Time	S	X
$t_1$	$S_1$	$X_1$
$t_2$	$S_2$	$X_2$
$t_3$	$S_3$	$X_3$
$t_4$	$S_4$	$X_4$
...	...	...



And then estimate a specific growth rate at each time...

Time	S	X	$\mu$
$t_1$	$S_1$	$X_1$	
$t_2$	$S_2$	$X_2$	$\mu_2$
$t_3$	$S_3$	$X_3$	$\mu_3$
$t_4$	$S_4$	$X_4$	$\mu_4$
...	...	...	

Recall...  $\frac{1}{X} \frac{dX}{dt} = \mu$

If time intervals are all equal, integrating (e.g.,  $t_1$  to  $t_3$ )...

$$X_3 = X_1 e^{\mu_2(t_3-t_1)}$$

So...  $\mu_2 = \frac{\ln(X_3) - \ln(X_1)}{t_3 - t_1}$

Method subject to significant error because S, X values are not always precise.

2) Fit one of the above models [ $\mu = f(S)$  or  $\mu = f(S, X)$ ]

Method doesn't work very well for  $K_S$  because its value is so small that  $\mu \approx \mu_{MAX}$  for the entire experiment.

...but, it is very easy to calculate  $\mu_{MAX}$  and to determine when lag, exponential and stationary phases occur.

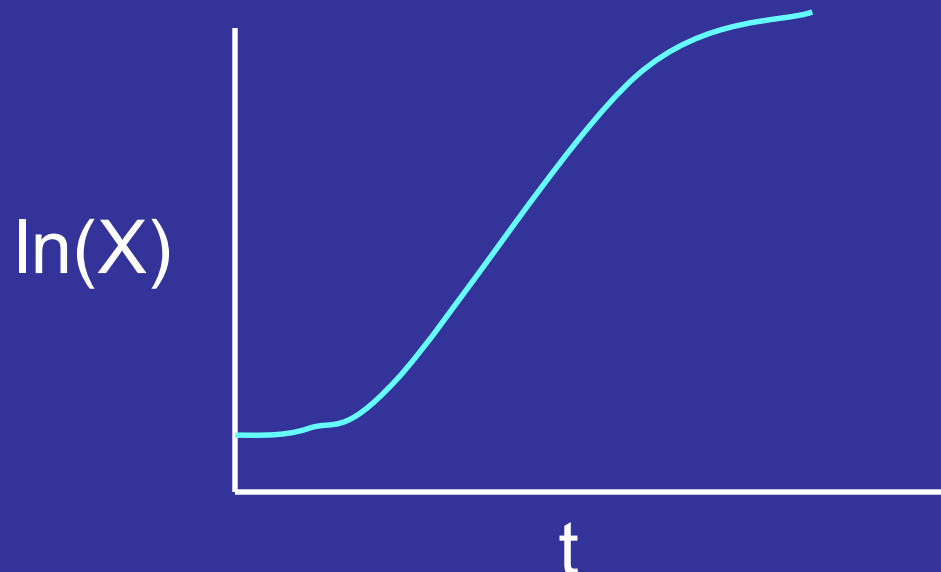
$$\frac{1}{X} \frac{dX}{dt} = \mu$$

Integrating...

(indefinite interval **during exponential growth**)

$$\ln(X) - C = \mu_{MAX}t$$

Plot of  $\ln(X)$  versus  $t$  will provide a slope of  $\mu_{MAX}$



## 2. Inhibitors

The presence of a chemical “inhibitor” (I) or substrate (S) or product (P) can reduce the cell growth rate. At sufficiently high concentration, cell growth ceases.

**Empirical** models are again used, and they have the general property:

$$\lim_{I \rightarrow \infty} \mu = 0$$

a) Substrate Inhibition  
Monod-like “Noncompetitive”

$$\mu = \frac{\mu_{\text{MAX}} S}{K_S + S} \frac{K_S'}{K_S' + S}$$

b) Substrate Inhibition  
Monod-like “Competitive”

$$\mu = \frac{\mu_{\text{MAX}} S}{K_S \left(1 + \frac{S}{K_I}\right) + S}$$

c) Product Inhibition  
Monod-like “Noncompetitive”

$$\mu = \frac{\mu_{MAX} S}{K_S + S} \frac{K_P}{K_P + P}$$

d) Product Inhibition  
Monod-like “Competitive”

$$\mu = \frac{\mu_{MAX} S}{K_S \left(1 + \frac{P}{K_P}\right) + S}$$

e) Product Inhibition  
Exponential

$$\mu = \frac{\mu_{MAX} e^{-\frac{P}{K_P}}}{\left(1 + \frac{K_S}{S}\right)}$$

f) Inhibition  
Monod-like “Noncompetitive”

$$\mu = \frac{\mu_{MAX} S}{K_S + S} \frac{K_I}{K_I + I}$$

g) Inhibition  
Monod-like “Competitive”

$$\mu = \frac{\mu_{MAX} S}{K_S \left(1 + \frac{I}{K_I}\right) + S}$$

Note that an inhibitor can be intentionally added to “force” a low growth rate, or to cause the substrate to be diverted to more product at the expense of growth.

### 3. Temperature

a) Ratkowsky Equation  
(J. Bacteriol. 1982, 149:1-5)

$$\mu^{1/2} = b(T - T_0) \quad \text{when } T > T_0$$

$$\mu^{1/2} = 0 \quad \text{when } T < T_0$$

This equation does not predict a maximum

Only useful for  $T < T_{\text{OPTIMAL GROWTH}}$

## b) Arrhenius Equation (like Enzyme Denaturation)

$$\frac{dX}{dt} = \mu X - \alpha X$$

Activation:  $\mu = A_a e^{-E_a/RT}$   $E_a = 18 \text{ kcal/mol}$

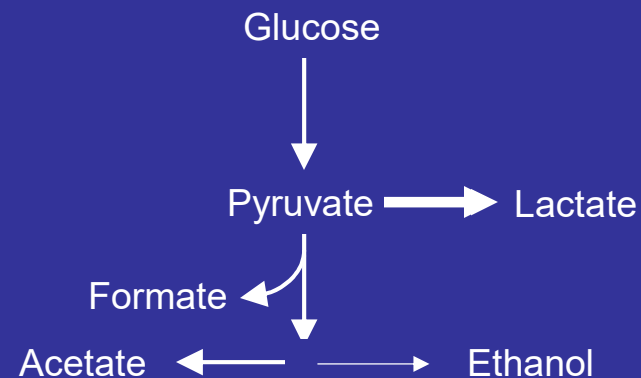
Death:  $\alpha = A_d e^{-E_d/RT}$   $E_d = 70 \text{ kcal/mol}$

## c) Other Comments

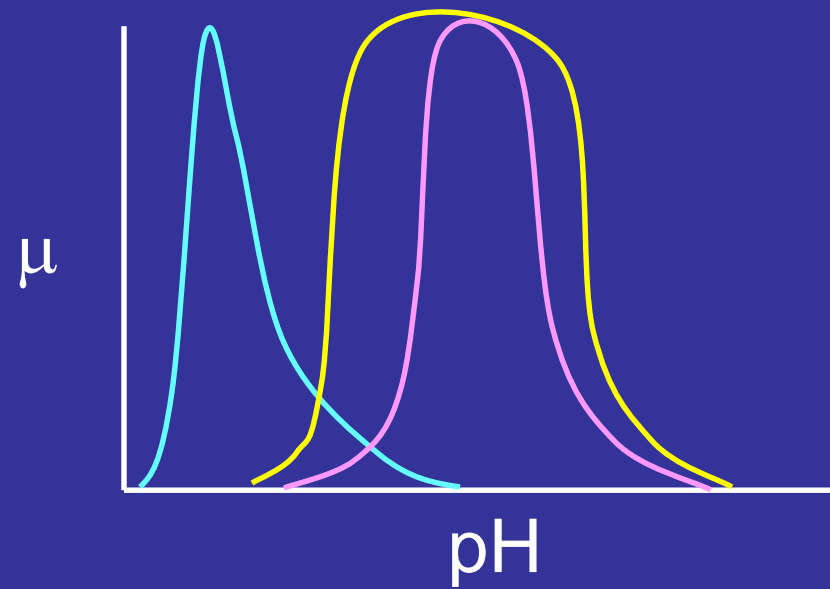
Temperature influences some enzymes more than others. Thus, the formation of product may be affected by temperature quite differently than cell growth. The consequence of this situation is:

$$Y_{P/S} = f(T)$$

Also,  $Y_{X/S}$ ,  $m_S = f(T)$



## 4. pH



Essentially impossible to predict

## 5. Logistic Equation

A different approach entirely

Specific growth rate is related to the **carrying capacity**

Carrying capacity ( $X_{\infty}$ ) is the maximum cell mass concentration that the environment can support.

$$\mu = k(1 - X / X_{\infty})$$

$$\frac{dX}{dt} = kX(1 - X / X_{\infty})$$

Integrating ( $X = X_0$  @  $t=0$ )...

$$X = \frac{X_0 e^{kt}}{1 - \frac{X_0}{X_{\infty}}(1 - e^{kt})}$$

## 6. Summary of Modeling Batch Growth – Unstructured Models

Growth

$$\frac{dX}{dt} = \mu X - \alpha X$$

Substrate Utilization

$$Q_S = m_S X + \frac{\mu X}{Y_{X/S}} + \frac{Q_P}{Y_{P/S}}$$

Product Formation

$$Q_P = Y_{P/X} \mu X \quad (\text{Growth-Associated})$$

Specific Growth Rate

$$\mu = \frac{\mu_{MAX} S}{K_S + S} \frac{K_P}{K_P + P}$$

# Volumetric Productivity

$$\Pi = P/t_{\text{FERM}}$$

